Basal Muscle Amino Acid Kinetics and Protein Synthesis in Healthy Young and Older Men

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ARCOPENIA, THE INVOLUNTARY decrease in muscle mass with aging, is associated with loss of strength and function, eventually leading to loss of independence.¹⁻⁴ The reduction in lean body mass also may contribute to the development of metabolic alterations, such as diabetes mellitus and osteoporosis.¹⁻⁵

The mechanisms leading to sarcopenia are still unclear. Numerous hypotheses have been suggested, including DNA damage,⁶ reduced protein synthesis,⁷ fiber type changes,² inactivity,⁸ inadequate nutrition,⁹,¹⁰ and hormonal changes.¹¹⁻¹⁴ A combination of several factors likely is responsible for the age-related changes in muscle mass and function.⁵ Overall, any of the primary factors responsible for sarcopenia should, at some point, affect muscle protein turnover by creating an imbalance between muscle protein synthesis and breakdown, thus inducing skeletal muscle loss. Therefore, a better understanding of the metabolic alterations characterizing sarcopenia may help to clarify the etiological characteristics of this condition and to determine the efficacy of potential treatments.

Initial studies on muscle protein metabolism have suggested that the metabolic alteration responsible for sarcopenia is a reduction in basal muscle protein synthesis rate in older people as compared with younger controls.¹⁵⁻¹⁹ However, more recent data did not confirm those results.²⁰⁻²² A careful review of these studies allows the exclusion of methodological problems as a reason for these discrepancies, since the methods used were similar and already validated in a variety of conditions.

Thus, 3 other possible explanations need to be considered. First, protein syn...
thesis alone does not provide enough information about muscle loss or gain because protein breakdown influences net muscle protein balance as well. Muscle protein breakdown was directly measured only in the 2 studies from our group,\textsuperscript{20,22} whereas the other studies did not measure breakdown\textsuperscript{16} or used indirect measures of breakdown (3-methylhistidine or creatinine urinary excretion),\textsuperscript{15,17-19,21} which are less sensitive and less specific than the methods used to measure protein synthesis, and they do not allow the estimation of muscle net protein balance. Second, the older population is more heterogeneous than the younger population, because the distinction between health and disease, activity and inactivity, or appropriate and inappropriate nutrition becomes less clear, thus increasing variability. Therefore, different investigators may have obtained contrasting results because they studied slightly different segments of the older population. Third, differences in the study design and insufficient power may have contributed to the discrepancies between different studies.

To assess if age is associated with a reduction in basal net muscle protein synthesis, we measured basal muscle protein turnover and amino acid kinetics in healthy young and older men, and we applied 3 different models to estimate muscle protein turnover to reduce a possible methodological bias. The volunteers were carefully selected to exclude any detectable confounders (eg, diseases, medications, exercise training) that might have affected measures of muscle protein turnover, and they were encouraged to maintain their usual lifestyle prior to the study to avoid the potential bias introduced by acute changes in diet, physical activity level, or both.

**METHODS**

**Subjects**

Forty-eight young and older men were recruited from June 1997 to July 2000 through the Sealy Center on Aging at the University of Texas Medical Branch (UTMB), Galveston, Tex (TABLE 1). There were 26 men in the young group with a mean (SE) age of 28 (2) years, and 22 men in the older group with a mean age of 70 (1) years. Women were not included because of difficulties in recruiting an adequate number representative of the population makeup (≤50% per sex). After explaining the purpose and risks of the study, which was approved by the institutional review board of UTMB, written consent was obtained from each volunteer. Four young and 4 older subjects had participated in a previous study.\textsuperscript{22} Volunteers were considered to be eligible if they were found to be healthy on the basis of clinical history, including a standard questionnaire on the activities of daily living (ADLs) in use at UTMB, physical examination, and screening tests, including complete blood count, blood chemistry workup, hepatitis panel, human immunodeficiency virus test, urinalysis, blood pressure, oral glucose tolerance test if necessary,\textsuperscript{23} and electrocardiogram.

Exclusion criteria were the following: cardiac, pulmonary, liver, or kidney disease; peripheral vascular disease; cerebrovascular disease; recurrent deep venous thrombosis; acute, subacute, or chronic infection; active cancer; autoimmune disease; metabolic disease; diabetes mellitus and glucose intolerance as defined by the American Diabetes Association\textsuperscript{21}; other endocrinopathies; anemia; hypertension treated with calcium channel blockers, angiotensin-converting enzyme inhibitors, \(\beta\)-blockers, or angiotensin receptor blockers; drug or alcohol abuse; coagulation defects; strength or aerobic training; impairment in the ADLs; history of falls; and anabolic steroid, corticosteroid, or antiandrogen therapy. Subjects with a history of anabolic steroid use or long-term corticosteroid therapy were excluded. Mild osteoarthritis was not an exclusion criterion, provided that it did not impair the ADLs. However, volunteers who could not discontinue analgesic or anti-inflammatory therapy for the time necessary for a complete washout (eg, 5 days for aspirin) were excluded. A careful clinical assessment of the amount of daily physical activity was performed to exclude any subjects engaged in regular aerobic or resistance exercise training, subjects whose jobs included daily heavy weight lifting or long walks, subjects with a history of falls (≥2/yr), or subjects with any impairment in ADLs. We did not use a standardized questionnaire to measure the level of physical activity because several studies have shown that the individual variability is high, and the validity of these questionnaires for the estimation of the daily total energy expenditure is limited.\textsuperscript{24,25}

The volunteers performed their regular activities and maintained their usual diet during the week preceding the study. This routine was preferred to an early admission to the General Clinical Research Center (GCRC) and to the administration of a standardized diet for the 3 to 5 days preceding the study, as previously reported,\textsuperscript{15-19,21} because these manipulations might influence basal muscle protein turnover.

**Table 1. Characteristics of the Subjects**

<table>
<thead>
<tr>
<th></th>
<th>Young Men</th>
<th>Older Men</th>
<th>(P) Value</th>
<th>Power ((\beta))</th>
<th>Least Significant Number†</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age, y</td>
<td>28 (2)</td>
<td>70 (1)</td>
<td>&lt;.001</td>
<td>1.00</td>
<td>4</td>
</tr>
<tr>
<td>Height, cm</td>
<td>176 (1)</td>
<td>175 (2)</td>
<td>.79</td>
<td>.06</td>
<td>5187</td>
</tr>
<tr>
<td>Weight, kg</td>
<td>76 (2)</td>
<td>80 (3)</td>
<td>.22</td>
<td>.23</td>
<td>249</td>
</tr>
<tr>
<td>Body mass index, kg/m²</td>
<td>24 (1)</td>
<td>26 (1)</td>
<td>.07</td>
<td>.44</td>
<td>114</td>
</tr>
<tr>
<td>Total leg volume, L</td>
<td>10.83 (0.43)</td>
<td>9.60 (0.32)</td>
<td>.03</td>
<td>.58</td>
<td>79</td>
</tr>
<tr>
<td>Leg blood flow, mL/min per 100 mL of leg volume</td>
<td>3.37 (0.25)</td>
<td>2.79 (0.20)</td>
<td>.08</td>
<td>.41</td>
<td>123</td>
</tr>
<tr>
<td>Total testosterone, nmol/L‡</td>
<td>21.26 (1.62)</td>
<td>19.66 (2.68)</td>
<td>.62</td>
<td>0.08</td>
<td>1262</td>
</tr>
</tbody>
</table>

*Values are the mean (SE).
†Total number of subjects (sum of both groups) that would produce an \(a = .05\) with \(\beta = .80\), and \(a\) and \(\beta\) equal to those of the data.
‡Measured in 19 young and 21 older men.
After the insertion of the femoral artery and venous catheters, L-[ring-2H5]-phenylalanine was infused for 5 hours. Indocyanine green (ICG) was infused from 230 to 270 minutes to measure rate of blood flow. At the time indicated by the arrows, muscle biopsy samples were taken and blood samples from femoral artery, femoral vein, and wrist vein were collected.

**Study Protocol**

The night before the study each subject was admitted to the GCRC. After 10 PM the subject was allowed only water ad libitum. At 6 AM on the morning of the study, a catheter was inserted in a forearm vein for phenylalanine tracer infusion and another catheter was inserted in the opposite forearm for blood sampling (Figure 1). Catheters were inserted in the femoral artery and femoral vein of one leg for blood sampling. The arterial catheter also was used for indocyanine green (ICG) (IC-Green, Akorn Inc, Buffalo Grove, Ill) infusion. Leg volume was measured using an anthropometric method. After the insertion of the femoral arteriovenous (AV) catheters and the collection of a blood sample for background phenylalanine enrichment and ICG concentration, a primed-continuous infusion of L-[ring-2H5]-phenylalanine (priming dose: 2 µmol/kg; infusion rate: 0.05 µmol·kg⁻¹·min⁻¹) was started at 6:30 AM (0 minutes) and continued for 5 hours. The background sample also was used to measure total testosterone concentration in 19 young and 21 older men. At 120 minutes, a first muscle biopsy sample was taken from the lateral aspect of the vastus lateralis of the leg with the femoral catheters, using a 5-mm Bergstrom needle. The muscle tissue sample (50-150 mg) was quickly blotted and frozen in liquid nitrogen and kept at −80°C until analysis. At 230 minutes, the continuous infusion of ICG was started in the femoral artery (0.5 mg/min) and carried out until 270 minutes. To measure ICG concentration, 4 blood samples were taken every 10 minutes from the femoral vein and the forearm vein during the infusion. After stopping the ICG infusion, to measure phenylalanine concentration and enrichment, 4 blood samples were taken every 10 minutes from the femoral artery and vein until 300 minutes. At 300 minutes, before stopping the tracer infusion, a second muscle biopsy sample was taken as described above.

Approximately 1 week after the study, leg muscle volume was measured using magnetic resonance imaging (MRI) (GE Signa 1.5 T whole body imager, General Electric, Milwaukee, Wis) in the first 10 older and 7 young subjects participating in the study. The remaining subjects could not undergo the procedure because the instrumentation was unavailable.

**Analytical Methods**

Muscle volume was calculated from the MRI scans as previously described. Indocyanine green concentration in infused and serum samples was measured spectrophotometrically at λ = 805 nm. Total testosterone was measured using a commercial radioimmunoassay (Diagnostic Products Corporation, Los Angeles, Calif). Phenylalanine concentrations and enrichments in blood samples were measured using gas chromatography mass spectrometry (GCMS; Hewlett Packard, Palo Alto, Calif). Muscle tissue from biopsy samples were processed as previously described. Free phenylalanine enrichment and concentration were measured after extraction from the muscle tissue samples using GCMS. Protein-bound phenylalanine enrichment was analyzed with GCMS after protein hydrolysis and amino acid extraction using the external standard curve approach for very low enrichments. The following 3 parameters are common to both the AV balance and the three-pool model:

1. Delivery to the Leg = Influx = $C_A \cdot BF$
2. Release from the Leg = Efflux = $C_V \cdot BF$
3. Net Balance = NB = $(C_A - C_V) \cdot BF$

The other kinetic parameters of the AV method were calculated as follows:

4. Rate of Appearance = Ra = $(C_A \cdot E_A / E_v) \cdot BF$
5. Release from the Muscle = $P = Ra - Influx = BF \cdot C_A \cdot [(E_v / E_A) - 1]$
6. Rate of Disappearance = Rd = $P + NB = BF \cdot [C_A \cdot (E_v / E_A) - C_V]$

**Calculations**

Muscle phenylalanine kinetic parameters were calculated using 2 different approaches: the AV balance method and the three-pool model recently described and validated. The AV balance method relies on the measurement of phenylalanine enrichments and concentrations in the femoral artery and vein to estimate muscle protein synthesis, breakdown, and net balance. These parameters are based on the extraction of labeled phenylalanine (tracer) from the femoral artery, the rate of appearance (Ra) of unlabelled phenylalanine (tracee) from the muscle in the femoral vein, and the net AV difference in phenylalanine concentrations, respectively. The three-pool model is an expansion of the AV method and relies not only on the measurement of phenylalanine enrichments and concentrations in the femoral artery and vein but also on the direct measurement of free phenylalanine enrichment in the tissue water. This method allows for the direct calculation of phenylalanine intracellular utilization for protein synthesis and release from protein breakdown as well as the calculation of the rates of phenylalanine transport from the artery into the tissue and from the tissue into the venous blood.
Eₐ and Eᵥ indicate phenylalanine enrichments (tracer/tracer ratio) in the femoral artery and vein, respectively. Data are presented per 100 mL of leg volume. In these calculations it is assumed that the phenylalanine enrichment in the vein is the closest to that of the intracelluar pool.

From the AV balance data, it is also possible to calculate the fractional uptake of labeled phenylalanine (%), which is another indirect index of muscle protein synthesis:

(7) Phenylalanine Fractional Uptake = \( \frac{(EₐCₐ - EᵥCᵥ)}{(EₐCₐ)} \)

The parameters specific to the three-pool model for labeled phenylalanine were calculated as follows:

(8) Transport Into Muscle = \( Fₐ,M = \frac{\left[ (Eₐ-Eᵥ)Cᵥ \right]}{(Eₐ)} \) BF

(9) Transport From Muscle = \( Fᵥ,M = \frac{\left[ (Eᵥ-Eₐ)Cₐ \right]}{(Eᵥ)} \) BF

(10) Arteriovenous Shunting = \( Fₐ,A = Influx- \frac{Fₐ,M}{Fₐ,A} \)

(11) Release From Proteolysis = \( Fₐ,M,0 = Fₐ,M \cdot \frac{(Eₐ/Eᵥ)}{-1} \)

(12) Utilization for Protein Synthesis = \( Fₐ,M = Fₐ,M,0 + NB \)

Eₐ indicates the phenylalanine enrichment (tracer/tracer ratio) in the muscle. Data are presented per 100 mL of leg volume.

Intracellular amino acid availability is given by the sum of transport into the muscle \( (Fₐ,M) \) and the muscle protein breakdown rate \( (Fₐ,M,0) \). Thus, it is possible to calculate protein synthesis efficiency as follows:

(13) Protein Synthesis Efficiency = \( \frac{Fₐ,M}{Fₐ,M+Fₐ,M,0} \)

Leg plasma flow was calculated at dye dilution steady state, as previously described. Leg blood flow was calculated by correcting the plasma flow by the hematocrit.

We also determined the fractional synthesis rate (FSR) of mixed muscle proteins by measuring the incorporation rate of L-[ring-2H₅]-phenylalanine into the proteins and using the precursor-product model to calculate the synthesis rate as follows:

(14) Fractional Synthesis Rate = \( \frac{\Delta Eₐ}{\frac{\left[ Eₐ(t₁)+Eₐ(t₂) \right]}{2}} \) 60-100

where \( \Delta Eₐ \) is the increment of protein-bound phenylalanine enrichment between 2 sequential biopsy samples, \( t \) is the time interval between the 2 sequential biopsy samples, and \( Eₐ(t₁) \) and \( Eₐ(t₂) \) are the phenylalanine enrichments (tracer/tracer ratio) in the free muscle pool in the 2 subsequent biopsy samples. The results are presented as percentage per hour.

We also calculated whole body phenylalanine Ra, an index of whole body protein breakdown, using the single-pool model:

(15) Whole Body Phenylalanine Ra = \( \frac{i}{Eₐ} \)

where \( i \) is the infusion rate of L-[ring-2H₅]-phenylalanine.

Statistical Analysis

Differences between young and older men were analyzed for each variable using the 2-tailed 2-sample \( t \) test. Since the prior studies reported contrasting results with regards to muscle protein synthesis, some observing a reduction with age and others finding no age-related differences, we could not calculate a priori the least significant number of subjects to be studied to reach a set significance with a set power. Thus, we opted for a large-scale study, relative to the technology used. On the basis of the results obtained, we calculated for each measured parameter the power and the least significant number of observations, which we defined as the total number of observations that would produce an \( \alpha = .05 \) with a \( \beta = .80 \), with \( \delta \) and \( \sigma \) equal to those of the data, to provide additional information regarding sample size for future investigations (JMP statistical software, version 4.0.2; SAS Institute Inc, Cary, NC). The relationships between total leg volume and leg muscle volume and between FSR and total testosterone concentration were assessed with the Pearson correlation coefficient. Significance was set at \( P < .05 \).

RESULTS

Subjects’ Physical Features

Height, weight, and body mass index were similar in the older and in the young men (Table 1). However, the total leg volume was significantly lower in the older than in the young men, suggesting a reduced leg muscle mass (Table 1). Mean (SE) leg muscle volume measured by MRI in 10 older and 7 young men was lower in the older than in the young men (young men, 6.16 [0.23] L; older men, 4.64 [0.26] L; \( P = .001 \)). There was a significant correlation between total leg volume and leg muscle volume (\( r^2 = .77; P < .001 \)) in the 17 subjects who underwent MRI measurement of leg muscle volume (Figure 2). Leg blood flow was not different between groups (Table 1).

Basal total testosterone concentration was measured in 19 young and 21 older subjects (Table 1). Total testosterone was slightly lower in the older than in the young men.

Phenylalanine Concentrations and Enrichments

Phenylalanine concentrations in the femoral artery and vein were significantly higher in the older than in the young men (\( P < .001 \)) (Table 2). However, no difference in muscle free phenylalanine concentration was found between groups.

Phenylalanine enrichments were slightly but significantly higher in the femoral artery (\( P = .04 \)) and in the muscle tissue water in the older men (\( P = .03 \)). Phenylalanine enrichment in the femoral vein was not different in the
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2 groups. The enrichments were at steady state during the sampling period (data available from authors).

Phenylalanine Kinetics
Phenylalanine delivery to the leg, release from the leg, and NB across the leg were similar in older and young men (Table 3). Using the AV balance method, phenylalanine Ra was not different in both groups. Muscle proteolysis was slightly but significantly higher in the older men (P = .045) (an N=91 would have been needed to achieve a power of β = .80). Phenylalanine AV shunting (FVAV) was slightly but significantly higher in the older men (P = .03), whereas the transport from the muscle (FVM) and the release from proteolysis (FVMn) did not differ in the 2 groups. Phenylalanine utilization for protein synthesis was significantly higher in the older group (P = .04), confirming the AV model data. Also, the fractional uptake of phenylalanine, another index of protein synthesis, was significantly higher in the older group (P < .001). Protein synthesis efficiency was similar in young and older men.

### Table 2. Phenylalanine Concentrations and Enrichments in Femoral Artery, Femoral Vein, and Muscle Tissue

<table>
<thead>
<tr>
<th>Phenylalanine</th>
<th>Young Men (n = 26)</th>
<th>Older Men (n = 22)</th>
<th>P Value</th>
<th>Power (β)</th>
<th>Least Significant No.†</th>
</tr>
</thead>
<tbody>
<tr>
<td>Concentration, µmol/L</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Artery</td>
<td>53 (2)</td>
<td>64 (2)</td>
<td>&lt; .001</td>
<td>.97</td>
<td>26</td>
</tr>
<tr>
<td>Vein</td>
<td>60 (2)</td>
<td>71 (2)</td>
<td>.001</td>
<td>.92</td>
<td>34</td>
</tr>
<tr>
<td>Muscle</td>
<td>115 (7)</td>
<td>125 (7)</td>
<td>.30</td>
<td>.18</td>
<td>348</td>
</tr>
<tr>
<td>Enrichment, tracer/tracee</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Artery</td>
<td>0.073 (0.002)</td>
<td>0.080 (0.003)</td>
<td>.04</td>
<td>.57</td>
<td>82</td>
</tr>
<tr>
<td>Vein</td>
<td>0.056 (0.001)</td>
<td>0.058 (0.002)</td>
<td>.38</td>
<td>.14</td>
<td>478</td>
</tr>
<tr>
<td>Muscle</td>
<td>0.042 (0.002)</td>
<td>0.048 (0.002)</td>
<td>.03</td>
<td>.58</td>
<td>80</td>
</tr>
</tbody>
</table>

*Values are the mean (SE).
†Total number of subjects (sum of both groups) that would produce an α = .05 with β = .80, and δ and σ equal to those of the data.

### Table 3. Leg and Muscle Phenylalanine Kinetics

<table>
<thead>
<tr>
<th></th>
<th>Young Men (n = 26)</th>
<th>Older Men (n = 22)</th>
<th>P Value</th>
<th>Power (β)</th>
<th>Least Significant No.†</th>
</tr>
</thead>
<tbody>
<tr>
<td>Common parameters, nmol/min per 100 mL of leg volume</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Delivery to the leg</td>
<td>182 (16)</td>
<td>175 (12)</td>
<td>.76</td>
<td>.06</td>
<td>4108</td>
</tr>
<tr>
<td>Release from the leg</td>
<td>202 (17)</td>
<td>194 (13)</td>
<td>.70</td>
<td>.07</td>
<td>2673</td>
</tr>
<tr>
<td>Net balance</td>
<td>−21 (2)</td>
<td>−19 (2)</td>
<td>.51</td>
<td>.10</td>
<td>890</td>
</tr>
<tr>
<td>AV balance model, nmol/min per 100 mL of leg volume</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Rate of appearance</td>
<td>234 (19)</td>
<td>241 (16)</td>
<td>.78</td>
<td>.06</td>
<td>4630</td>
</tr>
<tr>
<td>Endogenous proteolysis</td>
<td>53 (4)</td>
<td>66 (5)</td>
<td>.05</td>
<td>.52</td>
<td>91</td>
</tr>
<tr>
<td>Rate of disappearance</td>
<td>32 (3)</td>
<td>48 (5)</td>
<td>.004</td>
<td>.84</td>
<td>43</td>
</tr>
<tr>
<td>Three-pool model, nmol/min per 100 mL of leg volume</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Transport to the muscle</td>
<td>90 (8)</td>
<td>115 (9)</td>
<td>.054</td>
<td>.49</td>
<td>99</td>
</tr>
<tr>
<td>Transport from the muscle</td>
<td>111 (9)</td>
<td>134 (10)</td>
<td>.10</td>
<td>.36</td>
<td>141</td>
</tr>
<tr>
<td>AV shunting</td>
<td>91 (10)</td>
<td>60 (9)</td>
<td>.03</td>
<td>.58</td>
<td>80</td>
</tr>
<tr>
<td>Release from proteolysis</td>
<td>64 (5)</td>
<td>76 (6)</td>
<td>.14</td>
<td>.31</td>
<td>168</td>
</tr>
<tr>
<td>Utilization for protein synthesis</td>
<td>43 (4)</td>
<td>58 (5)</td>
<td>.04</td>
<td>.55</td>
<td>85</td>
</tr>
<tr>
<td>Mixed muscle protein FSR, %/h</td>
<td>0.0578 (0.0047)</td>
<td>0.0601 (0.0046)</td>
<td>.73</td>
<td>.06</td>
<td>3208</td>
</tr>
<tr>
<td>Other indexes of protein synthesis</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Phenylalanine fractional uptake</td>
<td>0.077 (0.006)</td>
<td>0.132 (0.015)</td>
<td>&lt; .001</td>
<td>.94</td>
<td>32</td>
</tr>
<tr>
<td>Protein synthesis efficiency</td>
<td>28 (2)</td>
<td>30 (2)</td>
<td>.37</td>
<td>.14</td>
<td>471</td>
</tr>
</tbody>
</table>

*AV indicates femoral arteriovenous. Values are the mean (SE).
†Total number of subjects (sum of both groups) that would produce an α = .05 with β = .80, and δ and σ equal to those of the data.

Mixed Muscle FSR
Mean (SE) mixed muscle FSR was not different in the older and in the young groups with a power of β = .06 (Table 3 and FIGURE 3). The FSR was 3.5% higher in the older group. We estimated that we should have studied at least 3208 subjects to achieve an α = .05, with β = .80 and σ equal to that of the present data. We calculated that 8 subjects per group would be required to observe a 30% difference in FSR between young and older men. No relationship was found between total testosterone concentration and FSR (FIGURE 4).

Whole Body Phenylalanine Ra
Mean (SE) whole body phenylalanine Ra, an index of whole body protein synthesis, was not different (P = .07) in the older group (0.64 [0.02] mmol·kg⁻¹·min⁻¹) and in the young group (0.70 [0.02] mmol·kg⁻¹·min⁻¹), with a power of β = .47. To achieve an α = .05 and a β = .80, 105 subjects should have been studied.

COMMENT
Contrary to the general notion that muscle protein synthesis is reduced with age, we found that the kinetic indicators of muscle protein synthesis (phenylalanine Rd, fractional uptake, utilization for protein synthesis, and mixed protein FSR) all tended toward the opposite direction: a higher protein synthesis rate in older men. Nevertheless, phenylalanine net balance...
ports volunteers were given a standardized diet for several days prior to the study.\textsuperscript{15-19,21} Recent data suggest that the recommended protein dietary allowance for people older than 55 years might be higher than that reported for adults younger than 55 years.\textsuperscript{30} which is the cohort in which dietary allowance was assessed. Thus, standardized diets may not be adequate for healthy older men who may be compensating in their everyday life with increased ingestion of protein. Also, we cannot exclude that the older muscle is more sensitive to inactivity, so that an early admission to the GCRC (3 days before the study) as described in previous studies\textsuperscript{15-18} might have contributed to a reduction in physical activity and, consequently, muscle protein turnover. This hypothesis is indirectly confirmed by the fact that 1 group, who found a slower muscle protein synthesis in older subjects after a 3-day admission to the GCRC,\textsuperscript{18} did not confirm those findings when studying the subjects the day after admission.\textsuperscript{21}

To assess the influence of age alone on muscle protein kinetics, we intentionally selected healthy and active, although not participating in exercise training, older subjects and compared them with healthy younger men. In addition, the older volunteers of our study had total testosterone concentrations within the mid- to normal range, and we found no difference in total testosterone concentrations between the young and older men. Since other investigators have found age-related differences in testosterone concentrations,\textsuperscript{11} the lack of a difference in testosterone concentration between the older and young men in our study might be explained by a high variability of total testosterone concentration in the older age group and by the fact that free testosterone was not measured. If our selection criteria allowed us to assess the effect of age per se on muscle protein turnover, our older volunteers may have not been representative of the general older population. Furthermore, it is possible that the subjects of other studies were not as healthy as the subjects in our study, although they were not clinically ill.

One question that our study cannot answer is whether age affects the turnover rates and the net balance of specific muscle proteins. Although the synthesis rate of isolated muscle proteins, such as myofibrillar proteins or, more specifically, myosin heavy chain, can be measured,\textsuperscript{15-17,19} at present no methods are available to determine their individual breakdown rates. However, myofibrillar proteins and myosin heavy chain represent the bulk of muscle proteins, and previous studies in which both mixed muscle proteins and muscle myosin heavy chain synthesis rates were measured together reported similar qualitative results.\textsuperscript{16,19} Thus, we be-
lieve that our data closely reflect contractile muscle protein turnover.

In conclusion, since differences in the basal rate of muscle protein turnover are not apparently responsible for the loss of muscle with aging, it follows that research on sarcopenia should focus on the response of muscle to specific stimuli, such as feeding and physical activity. Recent data indicate that a blunted response to feeding may be in part responsible for the slow loss of muscle with aging and that inactivity may be responsible as well, since exercise training results in increased muscle mass and muscle protein synthesis rate in both healthy and frail older individuals. Furthermore, age-related changes in the hormonal pattern and the possible pharmacological correction of these imbalances should be considered. Specifically, the reduction in sex hormones and growth hormone/insulin-like growth factor I in some subjects are appealing targets for a replacement therapy, which could positively impact sarcopenia. From a clinical perspective, it is encouraging that age-related reduction of basal muscle protein synthesis does not appear to explain sarcopenia, as it would be difficult to target such a fundamental response with an appropriate therapeutic intervention. On the other hand, it may be more feasible to treat alterations stemming from inactivity, altered response to nutrients, or mixed muscle protein synthesis rates in 78-84 and 23-32 yr olds. Am J Physiol Endocrinol Metab. 2000;278:E620-E626.

normalities of any of those components can interfere with complete bladder emptying. We report the case of a patient with symptomatic abdominal distension, which was found to be due to urinary retention with bilateral hydroureter. We believe that this was induced by chronic alcoholism.

Report of a Case. A 45-year-old man presented with progressive abdominal distension for 6 months as well as dysuria and urinary frequency for 2 months. His medical history was significant for chronic alcohol abuse of 20 years (10 beers per day). He denied dizziness, dyspnea, abdominal pain, nausea, vomiting, or tenesmus.

Physical examination revealed normal vital signs without orthostatic changes. The sclerae were anicteric. The abdomen was extensively distended but soft and nontender without shifting dullness. Bowel sounds could not be heard. The liver was 5 cm in the right middle clavicular line. There was no splenomegaly. The prostate was small and smooth, and the stool was hem negative. The patient had no paresthesia, weakness, or other neurological deficits. Laboratory data revealed normal electrolytes and blood cell counts except for an elevated mean corpuscular volume of 105.8 fL. Liver function tests, including prothrombin time, were normal except for aspartate transaminase of 129 U/L. Serum ethanol level was 244 mg/dL.

The results of an abdominal ultrasound suggested ascites and bilateral severe hydroureter. An abdominal computed tomography scan (Figure) showed bilateral hydroureretonephrosis with cortical thinning and marked distension of the urinary bladder, extending to the upper abdomen with compression of both ureters in the upper pelvis. No pelvic mass, adenopathy, or ascites was identified. The prostate and seminal vesicles were not enlarged.

A Foley catheter was inserted and 18650 mL of urine was drained over a 12-hour period. The patient reported complete relief from his abdominal distension. Cystoscopy confirmed bladder distension, but did not show any urethral obstruction. A suprapubic bladder tube was placed for recovery of bladder contractions. The patient was discharged and lost to follow-up.

Comment. Abdominal distension in the presence of alcoholism usually suggests ascites from portal hypertension. This patient’s abdominal distension, however, was caused by urinary retention possibly due to chronic alcohol-induced neuropathy. Chronic alcoholism can cause both central and peripheral neuropathy. Two cases of alcohol-induced bladder dysfunction have been reported previously. In 1 case, the etiology was thought to be thiamine deficiency, and alcoholic myelopathy in the other. Urinary retention as the presentation of alcoholism has only been reported once, but alcohol-induced urinary distension with bilateral hydroureret, as described here, has not been reported to our knowledge. Several pieces of evidence suggest that this patient’s urinary retention was due to alcoholic neuropathy. First, he had no symptoms or signs of either thiamine deficiency or alcoholic myelopathy. Second, neither physical examination, cystoscopy, nor imaging studies showed evidence of obstruction. Third, he had no medical conditions and was taking no medications or other drugs that would predispose him to urinary retention. Thus, we believe that this patient’s abdominal distension was caused by urinary retention due to neuropenic bladder with autonomic dysfunction, which in turn was secondary to chronic alcohol abuse.

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CORRECTION

Incorrect Regression Equation: In the Clinical Investigation titled “Basal Muscle Amino Acid Kinetics and Protein Synthesis in Healthy Young and Older Men” published in the September 12, 2001, issue of THE JOURNAL (2001;286:1206-1212), the regression equation in Figure 4 is incorrect. The intercept value should be 0.073292 (positive not negative value). The equation should thus be $y = -0.0006x + 0.073292$. 

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